

Introduction

Sample preparation is one of the essential steps of bottom-up proteomics. The PreOmics iST sample preparation kit is designed to assist researchers achieving best results with few sample preparation steps and little hands-on time. For sample-specific protocols and optimization visit www.preomics.com/downloads or contact info@preomics.com.

Kit Contents

The kit contains everything to perform a complete sample preparation. It includes all chemicals to denature, reduce and alkylate proteins, as well as the enzymes to perform a tryptic digestion and a final peptide cleanup.

| Component | Cap | Quantity | Buffer Properties | | | | Description | Storage |
|------------|-----|----------|-------------------|--------|-------|----------|---|---------|
| | | | Organic | Acidic | Basic | Volatile | | |
| DIGEST | ● | 2x | | | | | Trypsin/LysC mix to digest proteins. | -20°C |
| RESUSPEND | ● | 1x 2 mL | | | | ● | Reconstitutes lyophilized proteolytic enzymes. | RT |
| LYSE | ● | 1x 1 mL | | | ● | | Denatures, reduces and alkylates proteins. | RT |
| STOP | ● | 1x 1 mL | ● | ● | | ● | Stops the enzymatic activity. | RT |
| WASH 0 | ● | 1x 2 mL | ● | ● | | ● | Cleans peptides from phytochemicals. | RT |
| WASH 1 | ● | 1x 2 mL | ● | ● | | ● | Cleans peptides from hydrophobic contaminants. | RT |
| WASH 2 | ● | 1x 2 mL | | ● | | ● | Cleans peptides from hydrophilic contaminants. | RT |
| ELUTE | ● | 1x 2 mL | ● | | ● | ● | Elutes the peptides from the cartridge. | RT |
| LC-LOAD | ○ | 1x 1 mL | | ● | | ● | Loads peptides on reversed-phase LC-MS column. | RT |
| CARTRIDGE | | 8x | | | | | Cartridge for 1 to 100 µg protein starting material. | RT |
| WASTE | | 8x | | | | | 2.0 mL tube for collecting waste after washing steps. | RT |
| COLLECTION | | 8x | | | | | 1.5 mL tube for collecting peptides after elution. | RT |
| ADAPTER | | 8x | | | | | Enables a cartridge to be placed into a tube. | RT |

Pre-Requisites

Common lab equipment is required for the sample preparation.

| Equipment | Quantity and Description |
|-------------------|---|
| PIPETTE | Careful sample handling and pipetting reduces contaminations and improves quantification. |
| SAMPLE | Cryomilled plant material (or other means of cryogenic grinding with liquid nitrogen). |
| HEATING BLOCK | Two heating blocks are recommended to support protein denaturation and digestion. |
| CENTRIFUGE | 1.5/2.0 mL reaction tube centrifuges are required for loading, washing and elution. |
| SONICATOR | If the sample contains DNA, shear it by sonication (e.g. Diagenode Bioruptor®). |
| VACUUM EVAPORATOR | Vacuum manifolds evaporate volatile buffers from the eluate before LC-MS. |
| ULTRASONIC BATH | Optional: can be used to resuspend peptides. |

Procedure



Method

1. LYSE

- 1.1. Transfer 1-100 µg protein from cryomilled plant material into a clean 1.5 mL microreaction LoBind tube.
- 1.2. Add 100 µL **LYSE** (●). Place sample in a HEATING BLOCK (95°C; 1,000 rpm; 10 min). **NOTE1**
- 1.3. Shear the sample in a SONICATOR (10 cycles; 30 sec ON/OFF).
- 1.4. Optional: Spin down droplets (RT; max. 300 rcf; 10 sec).

2. DIGEST

- 2.1. Add 210 µL **RESUSPEND** (●) to **DIGEST** (●) (1 tube for 4 reactions), shake (RT; 500 rpm; 10 min), pipette up/down.
- 2.2. Add 50 µL **DIGEST** (●) to tube and place it in a pre-heated HEATING BLOCK (37°C; 500 rpm; 3 hours).
- 2.3. Add 100 µL **STOP** (●) to tube (precipitation may occur), shake (RT; 500 rpm; 1 min/pipette up/down). **SP**
- 2.4. Spin sample in CENTRIFUGE (16,000 rcf; 1 min).

3. PURIFY

- 3.1. Use **ADAPTER** to place **CARTRIDGE** in **WASTE** tube. Label all tubes.
- 3.2. Transfer supernatant from 2.4. to **CARTRIDGE**. Be careful not to damage the bottom layer of **CARTRIDGE**.
- 3.3. Spin **CARTRIDGE** in a CENTRIFUGE (3,800 rcf; 1-3 min). If needed, adjust time to ensure complete flow-through.
- 3.4. Add 200 µL **WASH 0** (●) to **CARTRIDGE**, repeat step 3.3.
- 3.5. Add 200 µL **WASH 1** (●) to **CARTRIDGE**, repeat step 3.3.
- 3.6. Add 200 µL **WASH 2** (●) to **CARTRIDGE**, repeat step 3.3. **SP**
- 3.7. Use **ADAPTER** to place **CARTRIDGE** in a fresh **COLLECTION** tube. Label all tubes.
- 3.8. Add 100 µL **ELUTE** (●) to **CARTRIDGE**, repeat step 3.7., keep flow-through in **COLLECTION** tube.
- 3.9. Repeat step 3.8., keep flow-through in the same **COLLECTION** tube.
- 3.10. Discard **CARTRIDGE** and place **COLLECTION** tube in a vacuum evaporator (45°C; until completely dry).
- 3.11. Add **LC-LOAD** (○) to **COLLECTION** tube. Aim for 1 g/L concentration (e.g. 100 µL to 100 µg protein starting material).
- 3.12. Sonicate **COLLECTION** tube in an ULTRASONIC BATH (5 min) or shake (RT; 500 rpm; 5 min). **SP**

NOTE1 Volumes of buffers can be adjusted according to protein starting amounts.

Lysis temperature should be between 60-95°C.

Visit our FAQ website for more information: www.preomics.com/faq.

SP - Storage Point:

At this point, close the peptide containing tube or **CARTRIDGE** using silicon lid.

Peptides can be frozen at -20°C. Storage of peptides should not exceed two weeks at -20°C.

For extended storage, finish the protocol and store at -80°C.

Data analysis

Consider the following as fixed modifications in your database search:

| MODIFICATION | DESCRIPTION | COMPOSITION | SPECIFICITY | MASS | UNIMOD # |
|--------------|-----------------------------|----------------------------------|-------------|-------|----------|
| ALKYLATION | Carbamidomethyl on cysteine | C ₂ H ₃ NO | [C] | +57Da | 4 |

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